

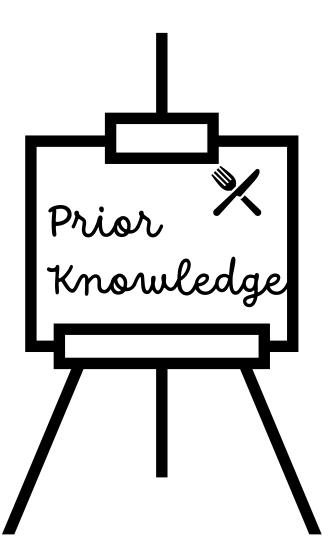
ViruSure GmbH

Impact of prior knowledge on viral clearance strategies

08th April 2024 Katy Lorineau, Head of GLP







1. What is it?

- 2. Where is it applicable?
- 3. How can it be used?



The notion *Prior knowledge* can already be found in ICH guidelines (e.g. Q2 (analytical procedure validation), Q8 (pharmaceutical development), Q10 (Pharmaceutical Quality System), Q14 (analytical procedure development)...

ICH Q5A (R2) definition:

Prior knowledge refers to existing knowledge and includes internal knowledge (e.g., development and manufacturing experience), external knowledge (e.g., scientific and technical publications, including vendors' data, literature, and peerreviewed publications), or the application of established scientific principles (e.g., chemistry, physics, and engineering principles).



Towards flexibility in viral clearance strategies

- Introduction of this new section offers possibility for new viral clearance strategies
- Annex 5 gives examples of process steps for which this concept can be applied
- Goals:
 - Encourage manufacturers to validation platform/in-house data
 - Reduce amount of product-specific virus clearance data
 - Save money
 - Save time
 - Save patients



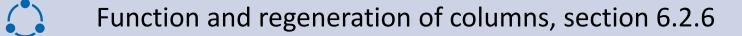
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Virus detection and identification: sections 3.2.1, 3.2.3





Replacement of spiked runs without product in mock solution or prior knowledge can be applicable, section 6.2.7



Reduction of number of steps based on previous virus clearance data, section 6.6 and Annex 5



Function and Regeneration of Columns

"Over time and after repeated use, the ability of chromatography resins and membranes used in the purification process to clear virus may vary." (section 6.2.6)

"Viral clearance studies should be performed to support media/resin reuse."

O.O. Ajayi et al.

Current Research in Biotechnology 4 (2022) 190-202

Table 3

Viral clearance database results for paired naïve and reused Protein A, AEX, and CEX chromatography resins against retrovirus, herpesvirus, reovirus, and parvovirus families.

Virus Family	Protein A			Anion Exchange			Cation Exchange		
	Number of paired records	Naïve resin LRV mean (log ₁₀)	Used resin LRV mean (log ₁₀)	Number of paired records	Naïve resin LRV mean (log ₁₀)	Used resin LRV mean (log ₁₀)	Number of paired records	Naïve resin LRV mean (log ₁₀)	Used resin LRV mean (log ₁₀)
Retrovirus	44	2.80	3.19	39	5.19	5.26	24	3.63	3.79
Herpesvirus	23	2.70	3.22	33	5.72	5.61	19	5.39	5.51
Reovirus	6	1.23	1.87	26	5.66	5.76	21	3.10	3.37
Parvovirus	14	2.08	2.55	35	4.64	4.75	17	1.62	1.68



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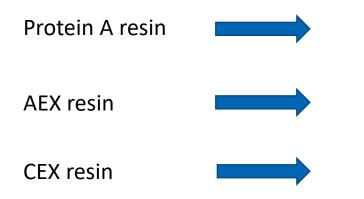
PDA Journal of Pharmaceutical Science and Technology

Retrospective Evaluation of Cycled Resin in Viral Clearance Studies—A Multiple Company Collaboration

John Mattila, Sherrie Curtis, Yenny Webb-Vargas, et al.

PDA J Pharm Sci and Tech 2019, 73 470-486 Access the most recent version at doi:10.5731/pdajpst.2018.009605

- Collaborative study for Protein A (97 paired observations) and AEX (144 paired observations) steps
- Evaluation of virus type, resin type, number of reuse cycles and virus challenge
- Clearance capability not impacted by resin cycling



Virus removal reported to be highly consistent for a given product Product-specific studies not expected

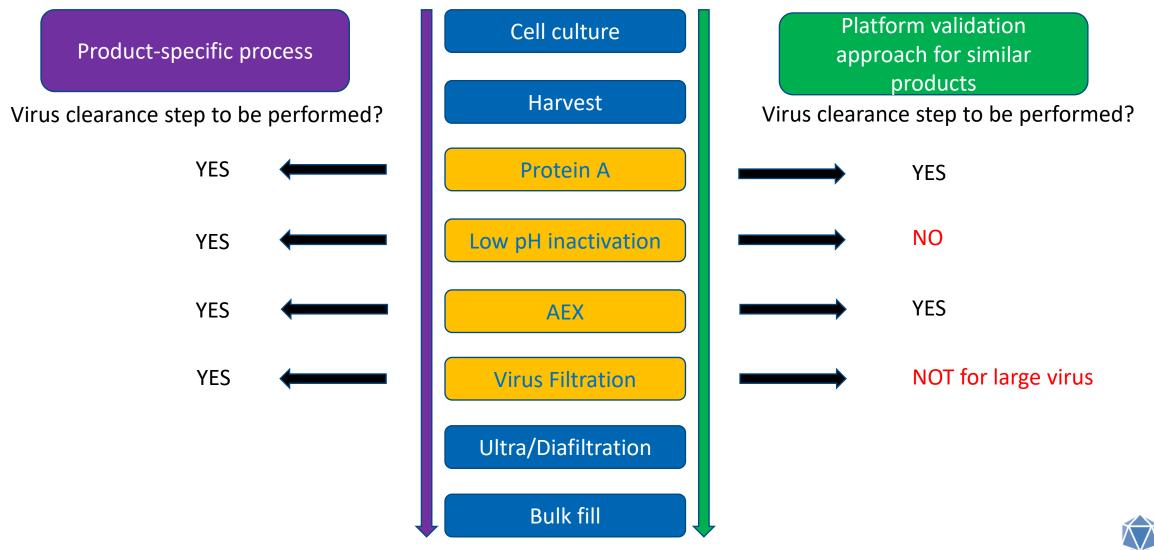
Product-specific studies not necessarily necessary In-house experience required and detailed justification

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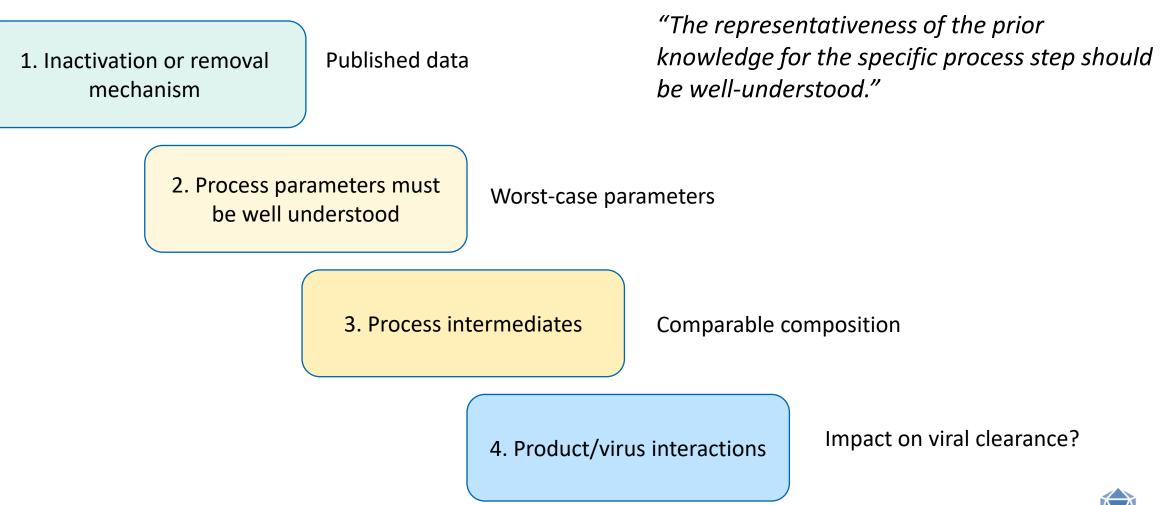
It must be assured that virus potentially retained by resin/media would be inactivated or removed before reuse.

Product-specific process validation vs. platform validation approach

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Annex 5: examples to reduce product-specific validation effort

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- **Platform validation** principles require robust viral clearance across products, established and wellcharacterised process conditions.
- Platform validation → New product viral clearance claims based on prior knowledge including inhouse experience:
 - Discussion of **all relevant platform data** available
 - **Rationale** to support platform validation approach
 - Comparison of new product and its process with other in-house products, process conditions and process intermediates
 - Process steps such as inactivation by detergent treatment, low pH and removal by virus filtration, **dedicated steps for viral clearance**, are suitable candidates for this approach.

Examples are only suggestions and should not be used as template or as sole basis for regulatory submission.



TRANSFUSION

BLOOD COMPONENTS

Robustness of solvent/detergent treatment of plasma derivatives: a data collection from Plasma Protein Therapeutics Association member companies

Herbert O. Dichtelmüller, Lothar Biesert, Fabrizio Fabbrizzi, Rodrigo Gajardo, Albrecht Gröner, Ilka von Hoegen 🔀 Juan I. Jorquera, Christoph Kempf, Thomas R. Kreil, Dominique Pifat, Wendy Osheroff, Gerhard Poelsler ... See fewer authors 🔿

First published: 26 August 2009 | https://doi.org/10.1111/j.1537-2995.2009.02222.x | Citations: 82

308 studies on plasma-derived products

RF between >2.9 and >6.5 log for the Triton X-100–TNBP combination

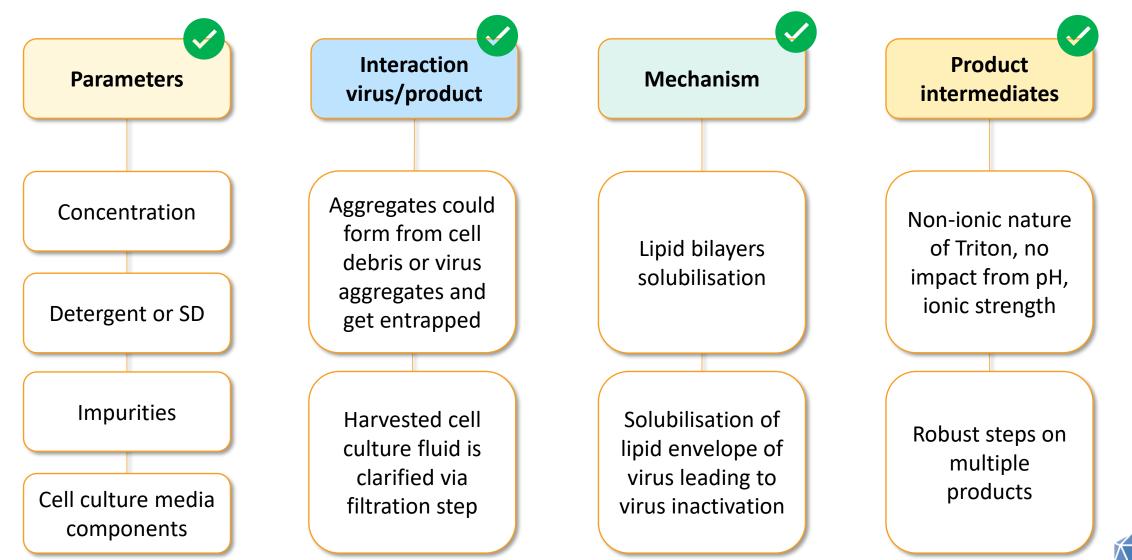
RF between >4.0 and >6.6 log for the same combination with polysorbate 80 added

ViruSure Internal Data on X-MuLV inactivation by Triton X-100 or Triton X-100 + TnBP						
Number of studies	10					
Triton concentration	0.3% to 1%					
Temperature	14-25°C					
Inactivation time	60-120 minutes					
Reduction factors	\geq 4.3 log to \geq 5.7 log					

ViruSure Data: 2010 to 2022



X-MuLV inactivation by Triton X-100 Treatment as Example 1



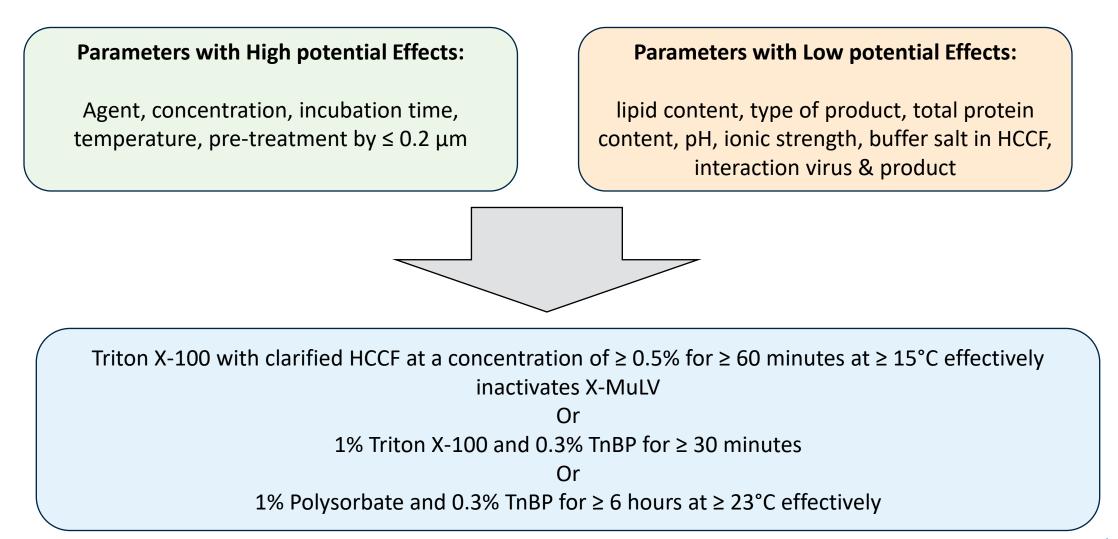
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S/D or Detergent treatment parameters and consensus







Bracketed Generic Inactivation of Rodent Retroviruses by Low pH Treatment for Monoclonal Antibodies and Recombinant Proteins

Kurt Brorson,¹ Sherrie Krejci,² Kitty Lee,² Elizabeth Hamilton,¹ Kathryn Stein,¹* Yuan Xu²

¹Division of Monoclonal Antibodies, Center for Biologics Evaluation and Research, Food and Drug Administration, 8800 Rockville Pike, Bethesda, Maryland 20892 ²Genentech, Inc., 1 DNA Way, South San Francisco, California 94080; e-mail: xu.yuan@gene.com

Received 17 July 2002; accepted 3 October 2002

- Low pH is a common step after Prot A for mAb purification
- Bracketed generic clearance: study defined the range of selected parameters which deliver consistently RF ≥ 4.6 log
- Performed on 1 product but can apply to others as long as key parameters are held constant
- The bracketing concept raised the possibility to define acceptable range which could be applied to all mAbs and Recombinant Proteins.

DOI: 10.1002/bit.28379 · Corpus ID: 257583099

Characterization of ionic strength for X-MuLV inactivation by low pH treatment for monoclonal antibody purification

Jena Daya, Valerie Cusick, J. Mattila • Published in Biotechnology and... 15 March 2023 • Medicine, Chemistry

TLDR Overall, robust and effective inactivation of X-MuLV at pH 3.65–3.80 can be achieved by manipulating either the pH or the NaCl concentration of theload material, and pH has a large effect when the load material has no additional NaCl.Expand

• pH is ≤3.8.

- Incubation time is ≥ 30 min.
- The incubation temperature is ≥14°C.
- The buffer system is acetate or citrate.
- Total protein concentration is ≤40 mg/mL.
- NaCl concentration is $\leq 500 \text{ mM}$.
- The p*I* of the mAb or RP is between approximately 3 and 9.





ViruSure Internal Data on X-MuLV inactivation by Low pHNumber of studies33 (out of 52)pH range3.2 – 3.85Temperature10-20°CInactivation time30-150 minutesReduction factors≥ 4.3 log to ≥ 6.5 log

ViruSure database: 2008 to 2023

Note:

19 studies with residual infectivity:

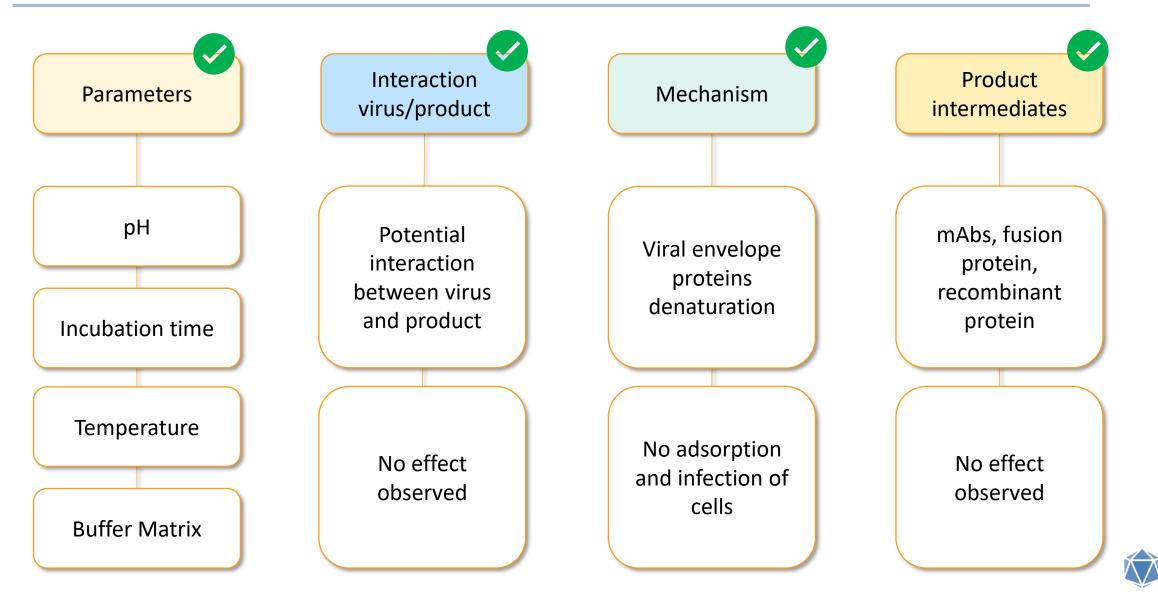
- pH too high (n= 13)
- Where < pH 3.6 (n= 6), very few positive wells detected in large volume plating testing, however RF remains high.



Low pH Treatment on X-MuLV as Example 2



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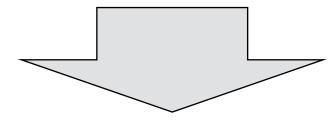


Parameters with High potential Effects:

pH, incubation time, temperature, buffer matrix

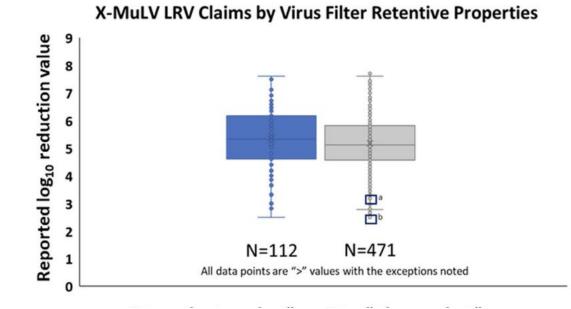
Parameters with Low potential Effects:

Product concentration, formation of protein aggregates, type of product, NaCl concentration (if less than 500 mmol/L)



Low pH at \leq pH 3.6, \geq 15°C for \geq 30 min at \leq 500 mmol/L NaCl Acetate buffer mostly used





Biotechnol Bioeng. 2022;119:743–761; Sarah A. Johnson

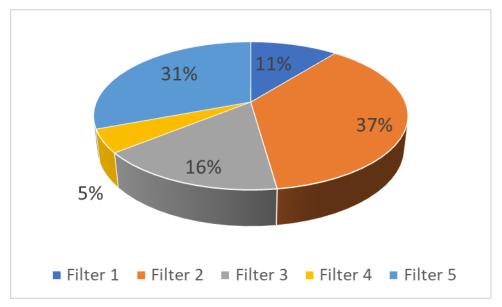
- FDA meta-analysis performed on the viral clearance data from monoclonal antibody (mAb) regulatory submissions from 1990 to 2015
- In all 112 large virus filter studies: complete removal
- In 469/471 studies with small virus filters: complete removal
- Detected breakthrough: no clear explanation if filter-related or study-related



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Virus Filters Tested*

* Based on data generated at ViruSure for studies between 2005 and 2018 (494 experiments)

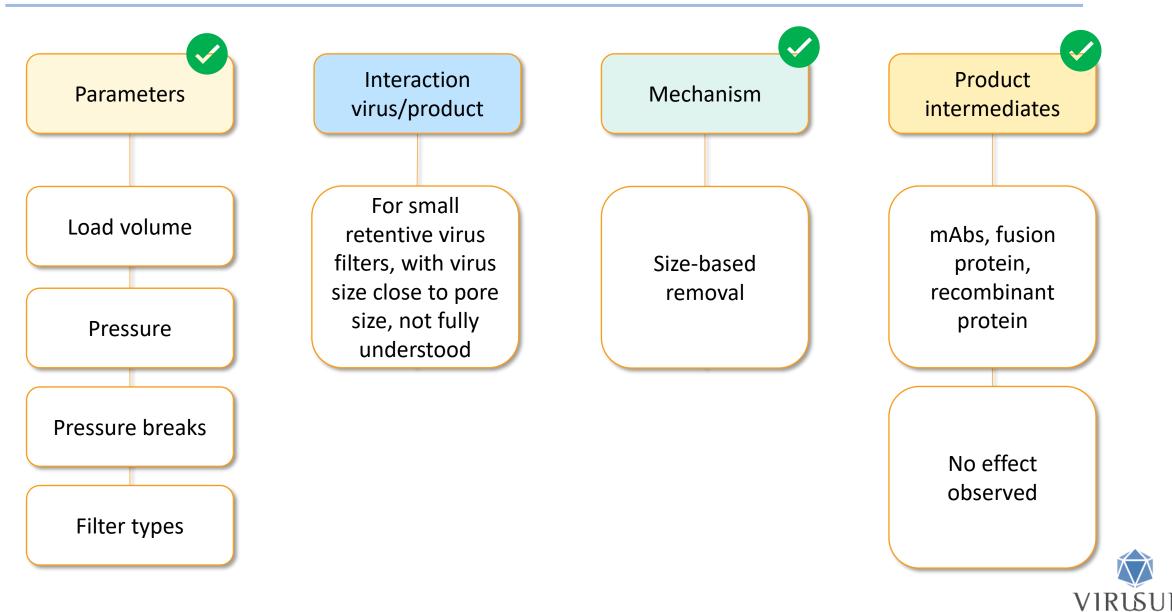
Filter	Total runs	Total runs with Residual infectivity				
Туре		PPV/MMV	Others (> size)			
Filter 1	53	18	0			
Filter 2	183	22	2			
Filter 3	81	1	1			
Filter 4	25	2	0			
Filter 5	152	6	2			

Filter 2: slight breakthrough for BVDV (1 pos well/(104)); 1 filter failure

Filter 3: slight breakthrough with X-MuLV (1 pos well/(104)) **Filter 5:** slight breakthrough with X-MuLV (1 pos well/(104) & 2(968) pos wells)



Virus Filtration: claim for large virus clearance as Example 3



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Virus Filtration parameters (using small virus retentive filters) and consensus Asahi

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Parameters with High potential Effects:

Volumetric throughput of load material, postwash, pressure/flow

Parameters with Low potential Effects:

Type of product, product concentration, pH, ionic strength, buffer matrix

Option 1: apply parvovirus RF to large viruses and envelopped viruses
BUT: underestimate virus removal in case of parvovirus passage (virus breakthrough)
Option 2: in-house data from both parvovirus and large viruses can build a plateform large viral clearance claim for commonly used small virus retentive filters

Thorough understanding of parameters with high effect should be in place.

If prior knowledge and in-house experience from other products used **to claim parvovirus removal**, at least one product-specific validation run (one confirmatory run) using parvovirus should be performed using worst-case conditions.

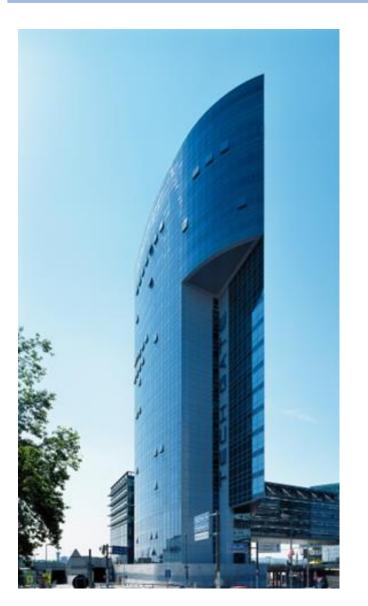


- ICH Q5A (R2) emphasizes the use of prior knowledge
- Possibility to decrease the number of validation runs to be performed
- In-house data must be demonstrated
- Annex 5 gives examples on how some steps can be used from a platform validation approach
- Other steps can follow this process assuming good external (large industry experience) and internal/in-house knowledge (data from similar products)



Thank you to the GLP team 🙂





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